

This is the post-print version of a paper published in Journal of Chromatography A

Citation for the original published paper (version of record):

Determination of adsorption isotherms in supercritical fluid chromatography. M. Enmark, P. Forssén, J. Samuelsson, T. Fornstedt. Journal of Chromatography A (2013) 1312, 124-133.

Access to the published version may require subscription.

doi:10.1016/j.chroma.2013.09.007

N.B. When citing this work, cite the original published paper.

**Attribution-NonCommercial-
NoDerivatives 4.0
International**



1 **Reproducibility between Modern Supercritical Fluid Chromatographic** 2 **Systems**

3 Martin Enmark, Jörgen Samuelsson, Patrik Forssén, Torgny Fornstedt*

4 Department of Engineering and Chemical Sciences, Karlstad University, SE-651 88 Karlstad, Sweden

5 *Corresponding author information:

6 Torgny Fornstedt, Professor, Karlstad University, SE-651 88 Karlstad, Sweden

7 Torgny.Fornstedt@kau.se, +46 54 700 1960; + 46 76 774 31 58

8 **Abstract**

9 This short communication is an investigation of adsorption isotherm estimation reproducibility between
10 different commercially available Supercritical Fluid Chromatography (SFC) instruments: Thar Super Pure
11 Discovery Series SFC, Waters UPC² and Agilent 1260 Infinity SFC. All experiments were conducted using
12 identical instrumental settings and identical column and the results clearly shows that analytical retention
13 times and “overloaded” elution profiles depends on both the instrument model and the particular instrument
14 of that model that is used. This means that dynamically determined adsorption isotherm will also depend on
15 the instrument, however it must be stressed that this do not mean that the adsorption is different. Instead it
16 means that identical instrumental settings will not reflect the conditions inside the column, most likely due to a
17 slight pressure-modifier difference between the systems despite of the identical operational settings.

18 This study highlights the importance of understanding the SFC-systems in order to to successfully transfer
19 method between systems, have reliable adsorption isotherm determination, to do analytical quality work and
20 to scale up from analytical to preparative mode. Additionally, despite the high complexity of SFC, the study
21 shows that the simplest column mass balance model, the ideal model assuming infinite column efficiency, can
22 be used to simulate elution profiles.

23 Keywords: Reproducibility; Supercritical fluid chromatography, overloaded elution profiles; modern SFC
24 systems

25 **1 Introduction**

26 Recently, both instrument manufacturers and the pharmaceutical industry have increased interest in
27 Supercritical Fluid Chromatography (SFC). The reasons for this growing interest are the lower environmental
28 impact and the considerably shorter separation times. Therefore a proper characterization and understanding
29 of the thermodynamics and kinetics of the adsorption processes is important, both for analytical and
30 preparative applications. Detailed thermodynamic information can be gained by conducting adsorption
31 isotherm measurements over a broad concentration range. If properly processed, these adsorption data gives
32 not only correct information about the degree of heterogeneity, but also the energy of interactions and
33 monolayer capacities of each individual type of adsorption site in the phase system [1,2]. We recommend the
34 following book and review of classical adsorption isotherms determination methods [3,4] and the recently
35 proposed work flow, and numerical tools, for collecting, processing and interpreting adsorption data [4–7].

36 In SFC, only a handful papers deals with the determination of adsorption isotherms in preparative
37 chromatography [8–12] and in even fewer cases the estimated adsorption isotherms have been verified by
38 comparing simulated and experimental overloaded elution profiles [8–10]. We suspect three reasons for this (i)
39 the lack of fundamental understanding of SFC, (ii) the lack of commercial SFC systems that has been properly
40 evaluated for fundamental studies and (iii) the relatively low interest for SFC during the past 20 years prior to
41 the recently growing interest. The lack of fundamental understanding of SFC is related to the state of the
42 mobile phase, i.e., its density and how it governs adsorption. Recently Tarafder published an article which
43 explains the origin of the variations of volumetric flow using a Thar system [13] and have quantified the density
44 and volumetric flow variations by measuring the mass flow. Also, models taking into account both heat- and
45 mass balance has recently been developed [14,15]. All of the above clearly motivates further development of
46 methods to determine and interpret adsorption isotherms as well as studies on how to properly simulate
47 elution profiles.

48 The investigation in this study is limited to methanol as modifier at operational conditions, temperature and
49 back pressure most typically observed when utilizing SFC to separate polar compounds [10]. Here we will use
50 the Elution by Characteristic Points (ECP) method [16–20] to determine the adsorption of antipyrine on neat
51 silica utilizing a fluid mixture of CO₂ and methanol. ECP requires that the separation system have high column
52 efficiency [17,18], which is often encountered in SFC, and in recent years the ECP method for liquid
53 chromatography (LC) has been considerably improved [16,19,20]. Because of this the ECP method is an
54 interesting candidate for SFC; still only a few reports exist on the use of the ECP method in SFC [21]. We will
55 perform identical experiments on a semi preparative Thar system and on analytical systems from Agilent and
56 Waters. The reason for choosing the two analytical systems from Agilent and Waters is that they represent two
57 of the latest commercial available systems while the semi-preparative instrument was added to widen the
58 study to include instruments used also for preparative purposes.

59 The first aim of this study is to investigate the reproducibility of adsorption isotherm determination between
60 different commercial available SFC systems and the second aim is to investigate if the ideal model can be used
61 to simulate elution profiles in SFC.

62 **2 Experimental Section**

63 Three different SFC systems were used in the study: a Thar Super Pure Discovery Series SFC (Waters
64 Corporation, Milford, MA, USA), an Agilent 1260 Infinity SFC system (Agilent Technologies, Palo Alto, CA, USA)
65 and a Waters UPC² system (Waters Corporation, Milford, MA, USA). Two Waters UPC² systems were used, one
66 placed at Karlstad University and one placed at Waters Corporation Saint-Quentin-en-Yveline (France). The
67 Thar system has only a circulating air oven with no dedicated heating of the incoming eluent, while both the
68 Agilent and Waters system have dedicated heaters prior to the column. The systems are hereafter referred to
69 as the “Thar”, “Agilent”, “Waters” and “Waters-Paris” system. Liquid CO₂ was supplied to the Thar/Waters
70 systems (AGA Gas AB, Sweden) and the Agilent system was fed gaseous CO₂. HPLC grade MeOH was used as

71 organic modifier and PT-100 resistance temperature detectors (Pentronic AB, Gunnebo, Sweden) connected to
72 a PT-104 data logger (Pico Technology, Cambridgeshire, United Kingdom) were used to measure the column
73 temperature.

74 A Kromasil neat silica 150×4.6 mm column, packed with 5 µm nominal particle size, 100 Å pore size (AkzoNobel
75 Eka, Bohus, Sweden) was used throughout the entire study. Antipyrine analytical standard was purchased from
76 Sigma-Aldrich (Sigma-Aldrich, Steinheim, Germany).

77 The chromatographic conditions: a back-pressure of 150 bar, a column temperature of 35°C and an eluent
78 composition of 85/15 (v/v) % CO₂ - MeOH. All experiments were carried out at volumetric flow of 2 mL/min. A
79 350 g/L stock solution of antipyrine dissolved in MeOH was used throughout the study. The obtained
80 chromatograms were then converted from UV-response to concentration by assuming a non-linear relation
81 between concentration and response [9].

82 The dead volume of the system capillaries and connections from the injector to the detector was estimated by
83 using injections of highly diluted samples of antipyrine when the column inlet and outlet capillaries were
84 connected using a union. The system dead volumes were estimated to be 190, 80 and 80 µL for the Thar,
85 Waters and Agilent system respectively.

86 Using a number of replicates of 5 µL injections of 0.35 g/L antipyrine the number of theoretical plates was
87 estimated to be approximately 9 500 on the Thar system and approximately 10 000 on the other systems.

88 **3 Results and Discussion**

89 First the reproducibility between different systems was studied by injecting 20 µL (full-loop) of 350 g/L
90 antipyrine on the four different SFC systems using identical instrumental settings and the same column. The
91 resulting overloaded elution profiles, corrected for system void volumes, is clearly system dependent as can be
92 seen in Fig. 1. This figure also shows that the elution volume is larger on the Thar system and the Waters and

93 Agilent system have lower, and similar, retention volumes. However the Waters-Paris system (same model as
94 the Waters system, see experimental section) had a retention volume that was more than 0.5 mL larger than
95 the Waters system when using the same column and the same set conditions. This indicates that either the set
96 conditions are not manifested in the systems or that the column conditions were different. Below we will
97 discuss and analyze the possible reasons for these differences prior to analyzing the estimated adsorption
98 isotherm and finally we will use the determined adsorption isotherms to predict elution profiles.

99 **3.1 Investigation of System Differences**

100 First we investigated if the volumetric flow rate was the reason for the observed difference between the
101 elution profiles shown in Fig 1. The column dead volume was determined and was estimated to be
102 approximately 1.9 mL on each system. This means that the volumetric flow rate was more or less identical on
103 all systems; otherwise the void volume would have been different. If the estimated void-volume is incorrect
104 this will lead to incorrect adsorption isotherm parameters and in the worst case even to the use of the wrong
105 adsorption isotherm model [22,23]. This issue is of major importance in a mechanistic investigation of the
106 adsorption. However, in this short study the main goal is to study differences in experimental elution profiles
107 between system and not to perform a mechanistic investigation.

108 Other important parameters governing retention are the density of the mobile phase, the fraction of organic
109 modifier and the column temperature. The density is a non-linear function of pressure and temperature [24]
110 and in order to calculate the density of the carbon dioxide and methanol stream inside the column, the
111 pressure at the column inlet and outlet, as well as the actual column temperature must be known. None of the
112 systems measures pressure directly prior and after the column, only at a point upstream the column and this
113 pressure reading cannot simply be correlated between different systems because the pressure drop upstream
114 to the back-pressure regulator will be divided between capillaries and column [25]. Pressure drop in the
115 capillaries will depend on their length and diameter, which cannot be assumed to be identical in all systems.

116 To investigate how the retention is affected by changes in system back pressure and modifier content we made
117 analytical injections, 5 μL of 0.350 g/L antipyrine, at back-pressures between 140 - 160 bar and modifier
118 fractions between 14 - 16 %, on the Waters system. As seen in Fig. 2 the retention volume is strongly affected
119 by changes in the modifier concentration and to a smaller extent to the set pressure. We noticed that the Thar
120 system has a much lower pressure drop over the system compared to the Waters and Agilent systems. This
121 sensitivity and back-pressure difference between systems are probably the explanation for the observed
122 difference in retention volume between the systems.

123 **3.2 Adsorption Isotherm Determination**

124 To determine adsorption isotherms using ECP it is advantageous if the volumetric flow rate is constant over the
125 column. At a set volumetric flow rate in an LC- instrument, both the mass flow and volumetric flow can be
126 assumed to be constant regardless of rather large pressure gradients across the column due to the
127 incompressibility of the solvents typically used. The mixture of 15 % MeOH and CO_2 is a compressible fluid,
128 albeit much less compressible than pure CO_2 . Because of this, the volumetric flow at any point in an SFC system
129 will be determined by the mass flow and density at that point. In this study, we will assume that we have
130 constant volumetric flow for both the simulations and adsorption isotherm determination. This assumption is
131 valid only if the inlet and outlet pressure are identical or such that, despite a non-zero pressure gradient, the
132 density gradient along the column is very small. We have also assumed that potential density gradients
133 throughout the column do not significantly affect the adsorption isotherm.

134 To validate the assumptions of constant volumetric flow, isothermal and iso-pycnic conditions, measurement
135 of the relative pressure drop and temperature profile of the column were performed on the Waters system. At
136 the column inlet and outlet PT-100 temperature probes were attached using a high strength and high thermally
137 conductive silver-epoxy adhesive. The recorded temperature was equal to 35°C with an uncertainty of 0.5°C. By
138 comparing the upstream pressure reading with the column installed and replaced by zero volume union, a

139 pressure gradient of 10 bars along the column was recorded at the studied conditions. This corresponds to inlet
140 and outlet densities of 871 and 867 kg/m³, a relative decrease of less than 0.5%, according to the REFPROP
141 software developed by National Institute of Standards and Technology. Also, by studying Fig. 2 in more detail,
142 the retention volume at 15 % (v/v) MeOH decreased from 6.6 to 6.5 mL, a relative decrease of 1.5 % for a
143 pressure drop increase from 150 to 160 bar. The indication of close to iso-pycnic and isothermal conditions
144 with constant volumetric flow through the column enables us to draw the conclusion that the approach of LC,
145 regarding determination of adsorption isotherms and simulation of elution profiles, is valid here, or at least a
146 good approximation.

147 To determine adsorption data, 20 µL injections of 350 g/L and 140 g/L, were done; see black lines in Fig. 3a and
148 Fig. 3b (note that both figures show the same experiments). The measurements repeatability was assured by
149 overlaying duplicated injections and by monitoring the back-pressure and temperature readings from the
150 system. It should be noted that the repeatability on the Thar system was worse than on the Agilent and Waters
151 systems. Only replicates with stable and low back-pressure ripple were used, which meant that much more
152 time had to be spent when using the Thar system. The adsorption isotherms were calculated from the 350 g/L
153 injection using the ECP approach, see Fig. 4. Prior to the model fit the adsorption data were analyzed using
154 Scatchard plots which were concave. This indicates that the adsorption describing the system is
155 heterogeneous. Tóth and bi-Langmuir models were fitted to the adsorption data and the bi-Langmuir model
156 was found to describe the data best. The ECP method requires that the separation system have high efficiency
157 and the determined efficiency for all systems were around 10 000 plates. Because all separations had high
158 efficiency, and only data from the injection with the highest loading were used for the adsorption
159 determination, the accuracy of this ECP can be expected to be good [18].

160 3.3 Simulation of Elution Profiles

161 Simulated elution profiles, using the determined adsorption isotherms and the Equilibrium-Dispersive (ED)
162 model [4], are plotted in Fig. 3a (gray lines). Since preparative SFC typically utilizes smaller particles, and the
163 diffusion is much faster than in liquid-based systems, it is reasonable to assume that the ideal model, to a much
164 larger extent than in LC, can accurately describe the column mass transfer phenomena, especially if the column
165 load is large [26]. To test this assumption, the same elution profiles were calculated using the ideal model and
166 compared with overlaid with experimental profiles, see Fig. 3b (gray lines). The agreement between the
167 experimental profiles and simulated profiles using both the ED and the ideal model were very good. The results
168 are not surprising because it is well established that the solution of the ideal model approaches that of the ED-
169 model when the Shirazi-number [26] increases. The Shirazi-number is a unit-less measurement of combined
170 column load and column efficiency and is approximately 70 for the 350 g/L injection which clearly indicates the
171 usefulness of the ideal model.

172 4 Conclusions

173 This study shows that the same instrumental set conditions does not necessarily give the same actual working
174 conditions over the column on different systems. In this study, recorded elution profiles on three different
175 systems, at the same set conditions, are clearly different. This means that it can be hard to transfer an
176 established separation method from one system to another, even if one is using the same column and identical
177 instrument settings. This has implications both for analytical methods and preparative applications, for
178 example different separation methods are usually screened on an analytical instrument and then the selected
179 method is transferred to a larger preparative system.

180 These issues can most likely be solved by measuring mass flow, pressure and temperature along the column,
181 together with an understanding of the density variations of the mobile phase [13,25]. We showed that in our
182 case the most probable explanation of the difference is that different system delivers different amount of

183 modifier (see Fig. 2) and that the column pressure is different. In other words: one SFC-system's 15% modifier
184 content is not the same as another system's 15% modifier content.

185 Despite the greater degree of complexity in SFC compared to LC, SFC separations generally have much higher
186 efficiency compared to the same separation conducted on an LC system. The increased efficiency, up to 10 000
187 theoretical plates in this case, enables us to use a much less complex mass balance model, the ideal model, see
188 Fig. 3, to calculate the elution profiles. This could be favorable for numerical optimization of SFC separations
189 where the mass balance model has to be solved thousands of times.

190 It should be noted that the investigation has focused on the case where one uses using carbon dioxide and
191 methanol mixtures as the mobile phase. The situation when using pure carbon dioxide, or other combinations
192 of polar co-solvents, remains to be investigated.

193 **Acknowledgements**

194 This work was supported by the Swedish Knowledge Foundation in the KK HÖG 2011 project "Improved
195 Purification Processes to Satisfy Modern Drug Quality Assurance and Environmental Criteria", by the Swedish
196 Research Council (VR) in the project grant "Fundamental studies on molecular interactions aimed at
197 preparative separations and biospecific measurements" and by the Research Council for Environment,
198 Agricultural Sciences and Spatial Planning for the project "High-Value Compounds from Agricultural and
199 Forestry Waste by Sustainable Methods, – an Interdisciplinary Approach for Bioresource Utilization".

200 **Figure captions**

201 **Fig. 1:** Experimental chromatograms recorded on the Thar (solid line), the Waters (dotted line), the Waters-
202 Paris (dash-dotted line) and the Agilent system SFC-system (dashed line). The set volumetric flow was 2 mL/min
203 and the injection volume was 20 µL of a 350 g/L sample.

204 **Fig. 2:** Retention volume of analytical peaks as a function MeOH content and pressure determined using 14,
205 14.5, 15, 15.5 and 16 (v/v) % MeOH content at 140, 145, 150, 155 and 160 bar on the Waters system. All
206 analytical injections were 5 μ L of a 0.35 g/L sample.

207 **Fig. 3:** Black lines are experimental chromatograms from the Thar (solid line), the Waters (dotted line) and the
208 Agilent system (dashed line). The corresponding gray lines are simulations in (a) using the equilibrium
209 dispersive model and in (b) using the ideal model. The set volumetric flow was 2 mL/min and the injection
210 volume was 20 μ L of a 350 g/L sample.

211 **Fig. 4:** Raw slope of the adsorption isotherm for all three systems determined using the ECP method: Thar
212 (solid line), Waters (dotted line) and Agilent (dashed line).

213 **References**

- 214 [1] J. Samuelsson, T. Fornstedt, *J. Chromatogr. A* 1203 (2008) 177.
- 215 [2] B.J. Stanley, G. Guiochon, *Langmuir* 10 (1994) 4278.
- 216 [3] A. Seidel-Morgenstern, *J. Chromatogr. A* 1037 (2004) 255.
- 217 [4] G. Guiochon, D.G. Shirazi, A. Felinger, A.M. Katti, *Fundamentals of Preparative and Nonlinear*
218 *Chromatography*, 2nd ed., Academic Press, Boston, MA, 2006.
- 219 [5] M. Enmark, J. Samuelsson, T. Undin, T. Fornstedt, *J. Chromatogr. A* 1218 (2011) 6688.
- 220 [6] J. Samuelsson, R. Arnell, T. Fornstedt, *J. Sep. Sci.* 32 (2009) 1491.
- 221 [7] F. Gritti, G. Guiochon, *J. Chromatogr. A* 1099 (2005) 1.
- 222 [8] A. Depta, T. Giese, M. Johannsen, G. Brunner, *J. Chromatogr. A* 865 (1999) 175.
- 223 [9] S. Ottiger, J. Kluge, A. Rajendran, M. Mazzotti, *J. Chromatogr. A* 1162 (2007) 74.
- 224 [10] C. Wenda, A. Rajendran, *J. Chromatogr. A* 1216 (2009) 8750.
- 225 [11] G. Brunner, M. Johannsen, *J. Supercrit. Fluids* 38 (2006) 181.
- 226 [12] M. Lübbert, G. Brunner, M. Johannsen, *J. Supercrit. Fluids* 42 (2007) 180.

- 227 [13] A. Tarafder, G. Guiochon, *J. Chromatogr. A* 1285 (2013) 148.
- 228 [14] K. Kaczmariski, D.P. Poe, G. Guiochon, *J. Chromatogr. A* 1217 (2010) 6578.
- 229 [15] K. Kaczmariski, D.P. Poe, G. Guiochon, *J. Chromatogr. A* 1218 (2011) 6531.
- 230 [16] J. Samuelsson, T. Undin, A. Törnecrona, T. Fornstedt, *J. Chromatogr. A* 1217 (2010) 7215.
- 231 [17] H. Guan, B.J. Stanley, G. Guiochon, *J. Chromatogr. A* 659 (1994) 27.
- 232 [18] L. Ravald, T. Fornstedt, *J. Chromatogr. A* 908 (2001) 111.
- 233 [19] J. Samuelsson, T. Undin, T. Fornstedt, *J. Chromatogr. A* 1218 (2011) 3737.
- 234 [20] J. Samuelsson, T. Fornstedt, *Anal. Chem.* 80 (2008) 7887.
- 235 [21] Y. Han, Y. Yang, P. Wu, *J. Chem. Eng. Data* 53 (2007) 16.
- 236 [22] J. Samuelsson, P. Sajonz, T. Fornstedt, *J. Chromatogr. A* 1189 (2008) 19.
- 237 [23] J. Samuelsson, J. Zang, A. Murunga, T. Fornstedt, P. Sajonz, *J. Chromatogr. A* 1194 (2008) 205.
- 238 [24] G. Guiochon, A. Tarafder, *J. Chromatogr. A* 1218 (2011) 1037.
- 239 [25] A. Rajendran, T.S. Gilkison, M. Mazzotti, *J. Sep. Sci.* 31 (2008) 1279.
- 240 [26] S. Golshan-Shirazi, G. Guiochon, *J. Chromatogr.* 506 (1990) 495.







